

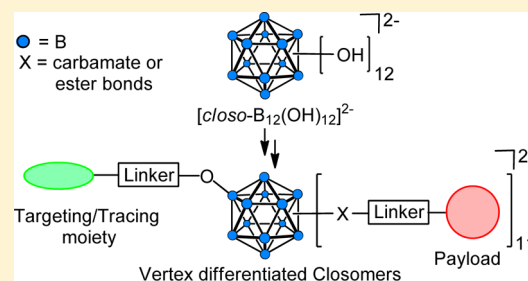
Synthesis of Vertex-Differentiated Icosahedral *closo*-Boranes: Polyfunctional Scaffolds for Targeted Drug Delivery

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S Supporting Information

ABSTRACT: We report methods for the synthesis of vertex-differentiated icosahedral *closo*-boranes. A single B–OH vertex of the icosahedral borane [*closo*-B₁₂(OH)₁₂]²⁻ was derivatized to prepare [*closo*-B₁₂(OR)(OH)₁₁]²⁻ using optimized alkylation conditions and purification procedures. Several representative vertex-differentiated icosahedral *closo*-boranes were prepared utilizing carbonate ester and azide–alkyne click chemistries on the surface of the *closo*-B₁₂²⁻ core.



Over the past decade, nanoscale carriers such as dendrimers, liposomes, and micelles have attracted much attention for their capacity to deliver multiple copies of therapeutic and diagnostic agents, potentially enhancing the clinical effectiveness of pharmaceutical agents. Nanocarriers can be constructed with or without the incorporation of a biological receptor-specific targeting moiety.^{1a} Nontargeted nanoscale pharmaceuticals have demonstrated improved specificity for tumor cells as a result of the “enhanced permeability and retention” effect.^{1b} The specificity of nanocarriers may be further increased by the addition of ligands (e.g., peptides, proteins, or antibodies) capable of selectively binding to receptors expressed only by target cells. Targeted nanocarriers are expected to be more effective and cause fewer side effects than conventional pharmaceuticals or nontargeted nanocarriers.

Polyhedral boranes have generated considerable interest in biomedical research in relation to their use as a ¹⁰B source in boron neutron capture therapy.² However, a still relatively unexplored application of polyhedral boranes is their use as nanocarriers. Our group has been actively pursuing research in this area and in particular is exploring the utility of derivatives of the icosahedral dodecahydro-*closo*-dodecaborate dianion [*closo*-B₁₂H₁₂]²⁻ such as [*closo*-B₁₂(OH)₁₂]²⁻, **1** (Figure 1), in the synthesis of discrete nanomolecular scaffolds for targeted, high-payload delivery of therapeutic and diagnostic agents.^{3,4}

The reactivity of the B–OH vertices in **1** resembles that of alcohols and can be used to anchor up to 12 radial arms with desired functionalities on the B₁₂²⁻ core. Previous reports from our laboratory have described syntheses of various 12-fold carbonate, carbamate, carboxylate ester, and ether derivatives, which are collectively referred to as “*closomers*”.⁵

An ideal nanocarrier for targeted drug delivery requires the presence of heterobifunctionalized linker arms within the same scaffold onto which one can install both a targeting moiety and

payload molecules by employing orthogonal chemistries. We envisage utilization of the known reactions of **1** to develop a variety of novel heterobifunctionalized, monodispersed closomers useful in development of novel nanoscale targeted drug delivery systems (Figure 1). Herein, we report methods for the synthesis of vertex-differentiated closomers that will permit the addition of both a targeting moiety and multiple payload molecules.

The first step in the synthesis of the vertex-differentiated closomers was reaction of a single BOH vertex in (TBA)₂-**1** (TBA = tetrabutylammonium) to produce the monoalkylated closomer **3** (Scheme 1). Key to the success of this study was to obtain **3** in pure form. Reaction optimization involved trying several stoichiometric ratios of (TBA)₂-**1** and the alkylating bifunctionalized linker **2**, employing different bases [*N,N*-diisopropylethylamine (DIPEA) and triethylamine (Et₃N)] and varying reaction temperature and time. Under the optimized etherification conditions of 0.7 equiv of **2** and 1.0 equiv of DIPEA in acetonitrile (ACN) at 85 °C for 24 h, a moderate yield of monoalkylated product **3** (36%) was achieved. On average, two to three size-exclusion chromatographic column sequences using lipophilic Sephadex LH-20 were necessary to completely separate **3** from the unreacted starting material **1** and minor amounts of multisubstituted products.

Purified product **3** was characterized by ¹H, ¹³C and ¹¹B NMR, HRMS and IR analysis. Whereas the ¹¹B NMR spectrum of **1** exhibits a sharp singlet at –18.24 ppm, that of **3** showed a broadened peak at –18.16 ppm consisting of all 12 B–O vertices, with broadening due to loss of symmetry in the structure (Figure 2). The IR-spectrum of **3** also displayed a

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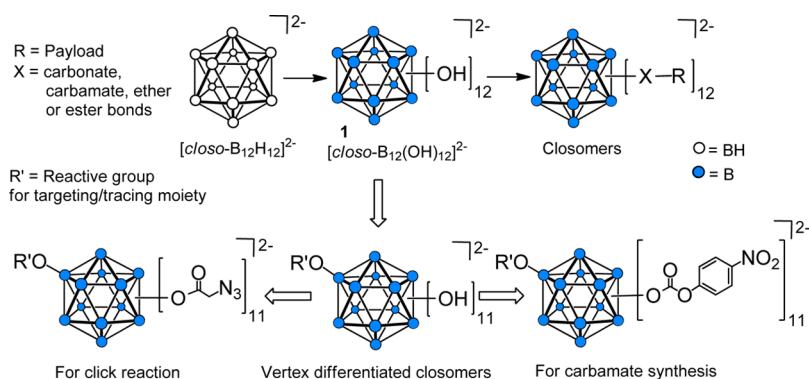
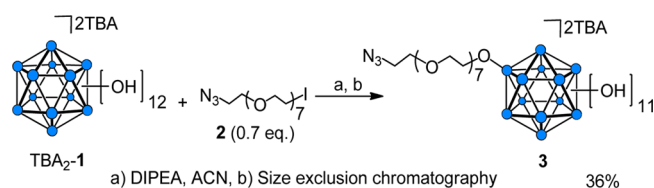


Figure 1. Schematic representation of organic synthesis on an icosahedral $closo-B_{12}^{2-}$ surface.

Scheme 1. Synthesis of Vertex-Differentiated Icosahedral $closo$ -Borane Scaffold 3



characteristic peak at 2106 cm^{-1} attributable to the asymmetric stretch of the azide group (Figure 3).

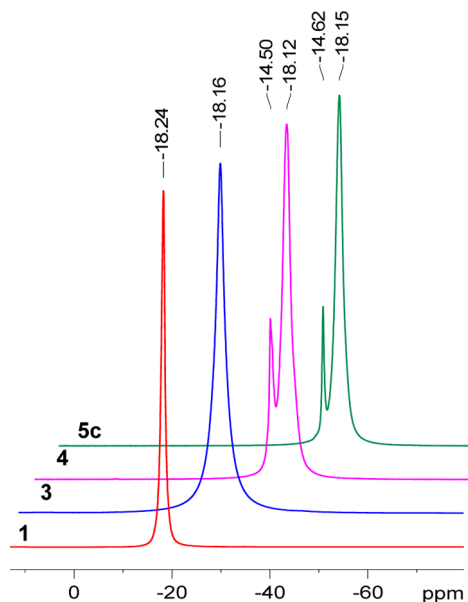


Figure 2. ^{11}B NMR (160.4 MHz, CD_3CN) stacks of closo-boranes **1**, **3**, **4**, and **5c** showing changes in chemical shifts as a result of substitution at the periphery of the cage.

For synthesis of **3**, choice of a suitable linker is crucial. The azido group at the distal terminus of **2** ($\text{N}_3\text{-OEG-I}$) is able to serve as a latent functional group during the multistep synthesis, and at a later stage it can be reacted with an alkyne-terminated ligand using a Cu(I) -catalyzed variant of the Huisgen 1,3-dipolar cycloaddition click reaction.⁶ The linker **2** was synthesized in four steps from octaethylene glycol (OEG).⁷

The remaining 11 hydroxyl groups on **3** were functionalized by employing closo-borane carbonate^{5c} ester reactions. Reaction of **3** with 4-nitrophenyl chloroformate (60.0 equiv) in ACN at 50

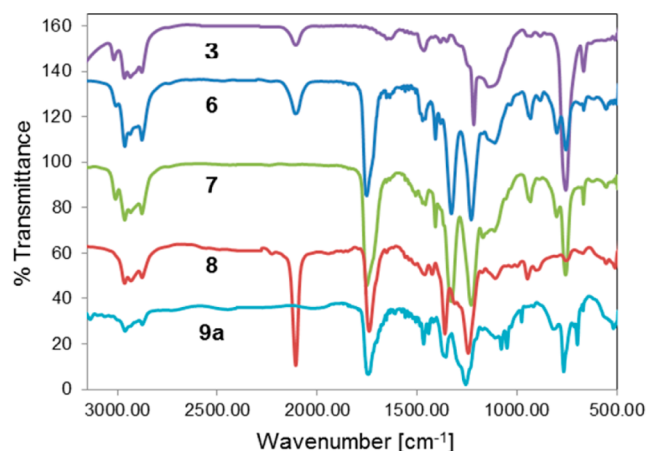


Figure 3. Characteristic changes in the IR spectra during synthesis of **9a**.

$^\circ\text{C}$ for 15 h produced the 11-fold 4-nitrophenyl carbonate (NPC)-substituted closo-borane **4** (Scheme 2). The yield was 63% following purification by size-exclusion chromatography on a lipophilic Sephadex LH-20 column. The purified closo-borane **4** was characterized by ^1H , ^{13}C , and ^{11}B NMR, HRMS, and IR analysis. The ^{11}B NMR spectrum of **4** showed two peaks, one at -14.50 ppm for the B-OR vertex and another at -18.12 ppm for the 11 B-OCOO- vertices (Figure 2). The IR spectrum of **4** also showed the presence of characteristic peaks at 2105 and 1766 cm^{-1} attributable to the asymmetric stretch of the azide and carbonate groups, respectively.

Reaction of 11-fold carbonate closo-borane **4** with various primary amines (5 equiv per vertex) produced the 11-fold carbamate closo-boranes **5a–c** in quantitative yields (Table 1). The purified closo-boranes **5a–c** were characterized by ^1H , ^{13}C , and ^{11}B NMR, HRMS, and IR analysis. The ^{11}B NMR spectra of **5c** contained two peaks, one at -14.62 ppm for the B-OR vertex, and one at -18.15 ppm for the 11 aggregate B-OC(=O)NH- vertices (Figure 2). The ^1H NMR spectra of closo-boranes **5a–c** lacked resonances in the aromatic region associated with the NPC group, indicating successful conversion of all carbonate groups.

Closo-borane **3** was also the starting material in a series of reactions that combined closo-borane ester^{5f} and click reactions to permit the generation of closo-boranes with a variety of functionalities. First, **3** was converted to the 11-fold chloroacetate closo-borane **6** (Scheme 3) by reaction with chloroacetic anhydride (4 equiv. per vertex). The yield was 86% after purification by size-exclusion chromatography on a

Scheme 2. Synthesis of Vertex-Differentiated Closomers via Carbonate Reactions

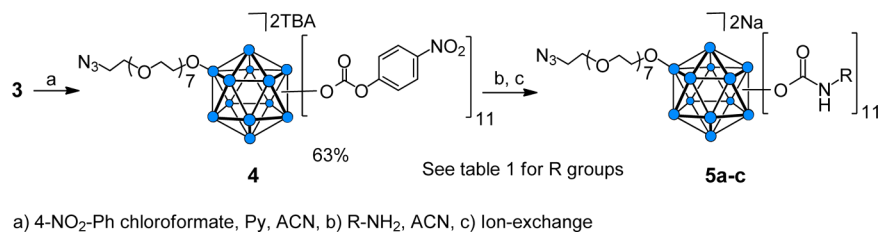


Table 1. Structure of Representative Amines and Yields of Closomers 5a–c

entry	R-NH ₂	yield (%)
5a	<i>n</i> -butylamine	92
5b	<i>N</i> -Boc-ethylenediamine	86
5c	<i>N</i> -Boc-2,2'-(ethylenedioxy)diethylamine	84

lipophilic Sephadex LH-20 column. The purified product **6** was characterized by ¹H, ¹³C and ¹¹B NMR, HRMS, and IR analysis. The ¹H NMR spectrum of **6** contained a characteristic peak at δ 4.1 ppm for the 22 protons assigned to the 11 Cl-CH₂CO₂- groups attached to the *closo*-B₁₂ cage. Similar to the ¹¹B NMR spectra of the closomer **5**, two peaks were seen in the ¹¹B NMR spectra of closomer **6**, one at -14.61 ppm for the B-OR vertex and one at -18.17 ppm for the 11 B-OCO vertices.

Next, a click reaction was employed to produce a Boc-protected amino-terminated linker from the lone azide group on closomer **6** (Scheme 3). Reaction of **6** with Boc-protected propargylamine in the presence of Cu(I) and DIPEA resulted in closomer **7** in high yield. ¹H NMR analysis of **7** showed a singlet at ~1.4 ppm for the nine protons specific to the Boc group. The absence of the characteristic asymmetric stretch of the azide group at 2107 cm⁻¹ from the IR spectrum of **7** confirmed the occurrence of the click reaction (Figure 3).

To permit addition of functionalities at the remaining 11 vertices of **7**, we employed our recently reported protocol^{Sf} to prepare the 11-fold azidoacetate closomer **8**. Reaction of closomer **7** with a 10-fold excess of sodium azide in dimethylformamide (DMF) at 50 °C for 2 days produced closomer **8** in quantitative yield (Scheme 3). The ¹¹B NMR spectrum of the azidoacetate closomer **8** did not change from that of the chloroacetate closomer **7**; however, the ¹H NMR spectrum showed that the characteristic peaks for the 22 protons assigned to the 11 Cl-CH₂CO₂- groups in closomer **7** shifted from δ 4.1 ppm to δ 3.7 ppm for the 11 N₃-CH₂CO₂-

groups in closomer **8**. The IR spectrum of **8** also exhibited a characteristic peak at 2106 cm⁻¹ attributed to the asymmetric stretch of the azide group (Figure 3).

The 11 azido groups of closomer **8** are excellent substrates for an azide-alkyne click reaction to attach payload molecules under mild reaction conditions (Scheme 3). Several representative aryl- and alkylalkynes (Table 2) readily reacted with **8** to

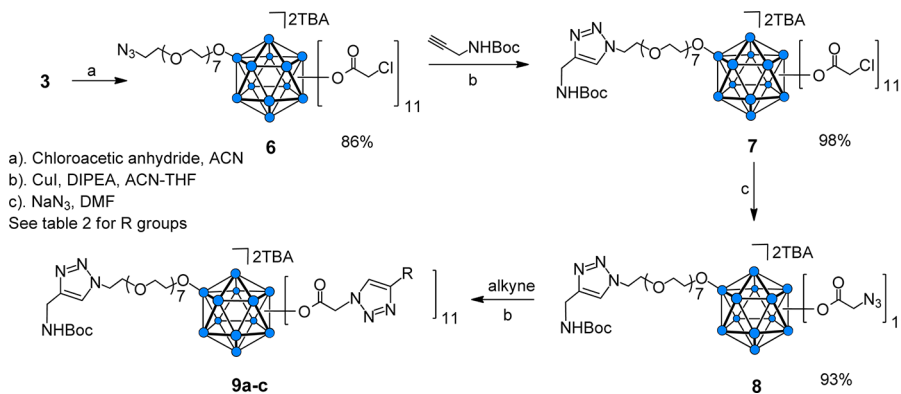
Table 2. Structures of Representative

entry	alkyne	yield (%)
9a	phenylacetylene	78
9b	1-hexyne	86
9c	propargyl acetate	81

produce the 11-fold triazole-linked closomers **9a–c** in excellent yields. All of the click products exhibited a characteristic singlet in the δ 7–8 ppm region in the ¹H NMR spectra for the 12 protons of alkene-CH of the 12 triazole rings of **9a–c**. The IR spectra of **9a–c** also showed loss of the characteristic peak at 2106 cm⁻¹ originally attributed to the asymmetric stretch of the azide groups in **8** (Figure 3).

In conclusion, we have developed methods for the synthesis of vertex-differentiated closomers possessing heterobifunctionalized linker arms. Optimized mono O-alkylation and appropriate purification methods permits conversion of a single hydroxyl group in [*closo*-B₁₂(OH)₁₂]²⁻ to the monoalkylated closomer [*closo*-B₁₂(O-OEG-N₃)(OH)₁₁]²⁻. The remaining 11 unreacted hydroxyl groups on the periphery of the icosahedral core can be readily transformed into highly reactive carbonate and azidoacetate closomers. The reactive end groups on each of the 11 linker arms allow for addition of suitable payload molecules via carbonate and click chemistry under very mild reaction conditions. The lone B-O-OEG vertex is spared and can be used for attachment of a targeting and or tracking moiety. This unique and versatile approach will be useful in the

Scheme 3. Synthesis of Vertex-Differentiated Closomers via Azide–Alkyne Click Reaction



construction of nanomolecular targeted delivery systems capable of carrying multiple copies of therapeutic or diagnostic imaging agents.

EXPERIMENTAL SECTION

General Information. Common reagents and chromatographic solvents were purchased from commercial suppliers and used without any further purification. NMR spectra were recorded on 400 and 500 MHz spectrometers. All NMR chemical shifts (δ) are reported in parts per million (ppm). The NMR peak assignments were confirmed using COSY experiments. The high-resolution mass spectrometry analysis was performed using ESI-TOF.

Synthesis of clesomer 3. A mixture of (TBA)₂-1 (1.15 g, 1.41 mmol) and ACN (40 mL) under stirring was allowed to reflux at 85 °C. To this mixture was added a solution of 2 (0.50 g, 0.99 mmol) and DIPEA (0.18 g, 1.41 mmol) in ACN (30 mL) slowly over 8 h at 85 °C. After the addition was completed, the reaction mixture was stirred for another 15 h at 85 °C, concentrated in the rotavap, and dried in vacuo to give the crude product as yellow oil. The product was then purified by size-exclusion column chromatography (lipophilic Sephadex LH-20) using methanol (MeOH) as the mobile phase. The product was obtained as a colorless oil. Yield: 0.43 g (36%). IR (neat): 3366, 2935, 2106, 1465, 1215, and 754 cm⁻¹. ¹H NMR (400 MHz, CD₃CN): δ 3.97 (m, 2H, -CH₂-PEG), 3.67–3.50 (m, 28H, -CH₂-PEG), 3.40 (t, 2H, *J* = 5.0 Hz, -CH₂-PEG), 3.12 (m, 16H, -CH₂-TBA⁺), 1.65 (m, 16H, -CH₂-TBA⁺), 1.39 (m, 16H, -CH₂-TBA⁺), 0.99 (t, *J* = 7.2 Hz, 24H, CH₃-TBA⁺). ¹³C NMR (100.6 MHz, CD₃CN): δ 71.1–70.8 (multiple peaks), 70.3, 59.2, 51.4, 24.2, 20.2, 13.6. ¹¹B NMR (160.4 MHz, CD₃CN): δ -18.16. HRMS (*m/z*): calcd for C₁₆H₄₃B₁₂N₃O₁₉ + TBA⁺ [M + TBA⁺]⁻ 953.6544, found 953.6191, calcd for C₁₆H₄₃B₁₂N₃O₁₉ [M]²⁻ 355.6841, found 355.6922.

Synthesis of Clesomer 4. A mixture of 4-nitrophenyl chloroformate (5.06 g, 25.1 mmol) and pyridine (2.15 g, 27.2 mmol) in ACN (50 mL) was stirred at 0 °C for 15 min. To this mixture, a solution of 3 (0.50 g, 0.42 mmol) in ACN (20 mL) was added slowly over 0.5 h at 0 °C. The reaction mixture was allowed to warm to rt and stirred for 15 h at 50 °C. The mixture was then concentrated to dryness, dissolved in dichloromethane (DCM), and filtered. The filtrate was concentrated and dried in vacuo, and the product was then purified by size-exclusion chromatography (lipophilic Sephadex LH-20) using ACN as the mobile phase. The product was obtained as a light yellow sticky solid. Yield: 0.80 g (63%). IR (neat): 3019, 2935, 2876, 2105, 1766, 1522, 1429, 1347, 1297, 1216, and 757 cm⁻¹. ¹H NMR (500 MHz, CD₃CN): δ 8.29 (m, 2H, -CH-PNPC), 7.41 (m, 2H, -CH-PNPC), 3.62–3.58 (m, 30H, -CH₂-PEG), 3.45 (m, 2H, -CH₂-PEG), 3.17 (m, 14H, -CH₂-TBA⁺), 1.69 (m, 14H, -CH₂-TBA⁺), 1.45 (m, 14H, -CH₂-TBA⁺), 1.05 (m, 21H, CH₃-TBA⁺) (lesser proton count for TBA⁺ is due to the exchange with H₃O⁺ and Na⁺). ¹³C NMR (125.7 MHz, CD₃CN): δ 156.7, 151.0, 145.2, 125.1, 122.4, 71.5–70.2 (multiple peaks), 58.3, 50.5, 24.8, 19.3, 12.8. ¹¹B NMR (160.4 MHz, CD₃CN): δ -14.50, -18.12. HRMS (*m/z*): calcd for C₉₃H₇₆B₁₂N₁₄O₆₃ [M]²⁻ 1264.2193, found 1264.2448.

General Procedure for the Synthesis of Carbamate Clesomer 5. A mixture of 4 (1 equiv) and amine (60 equiv) in ACN was stirred at 65 °C for 2–3 days. Progress of the reaction was monitored via mass spectrometry and ¹¹B NMR analysis. After completion, the reaction mixture was concentrated in a rotavap and dried in vacuo to obtain the crude product. The product was then purified by size-exclusion chromatography (lipophilic Sephadex LH-20) using MeOH as the mobile phase. Purified product was passed through an ion-exchange resin column to exchange TBA⁺ with Na⁺ ion.

Synthesis of Carbamate Clesomer 5a. Clesomer 5a was prepared from 4 (0.10 g, 0.03 mmol) and *n*-butylamine (0.15 g, 2.00 mmol) in ACN (10 mL) using the aforementioned general procedure for carbamate synthesis. The product was obtained as a colorless oil. Yield: 55.0 mg (92%). IR (neat): 3351, 2959, 2932, 2873, 2105, 1694, 1540, 1466, 1256, 1216, and 756 cm⁻¹. ¹H NMR (400 MHz,

CD₃CN): δ 6.40–6.33 (bs, 11H, NH), 4.01 (m, 2H, -CH₂-PEG), 3.68–3.53 (m, 28H, -CH₂-PEG), 3.39 (t, 2H, *J* = 5.2 Hz, -CH₂-PEG), 3.05 (m, 22H, -CH₂-butyl), 1.48 (m, 22H, -CH₂-butyl), 1.36 (m, 22H, -CH₂-butyl), 0.92 (t, 33H, *J* = 7.2 Hz, CH₃-butyl). ¹³C NMR (100.6 MHz, CD₃CN): δ 157.1, 72.8, 71.3–71.2 (multiple peaks), 65.9, 51.6, 42.7, 42.0, 32.7, 20.9, 14.3. ¹¹B NMR (160.4 MHz, CD₃CN): δ -16.31, -18.25. HRMS (*m/z*): calcd for C₇₁H₁₄₃B₁₂N₁₄O₃₀ + Na [M + Na]⁻ 1825.1121, found 1825.1139.

Synthesis of Carbamate 5b. Clesomer 5b was prepared from 4 (0.10 g, 0.03 mmol) and *N*-Boc-ethylenediamine (0.32 g, 2.00 mmol) in ACN (15 mL) using the aforementioned general procedure for carbamate synthesis. The product was obtained as a colorless oil. Yield: 80.0 mg (86%). IR (neat): 3353, 3017, 2978, 2108, 1698, 1516, 1216, and 758 cm⁻¹. ¹H NMR (400 MHz, CD₃CN): δ 6.15 (bs, 22H, NH), 4.08 (m, 2H, -CH₂-PEG), 3.66–3.59 (m, 28H, -CH₂-PEG), 3.39 (t, 2H, *J* = 5.2 Hz, -CH₂-PEG), 3.16 (m, 44H, -CH₂-ethylenediamine), 1.42 (s, 99H, -Boc). ¹³C NMR (100.6 MHz, CD₃CN): δ 157.5, 79.4, 71.2–70.6 (multiple peaks), 51.6, 42.3, 41.1, 28.6. ¹¹B NMR (160.4 MHz, CD₃CN): δ -16.31, -18.25. HRMS (*m/z*): calcd for C₁₀₄H₁₉₇B₁₂N₂₅O₅₂ [M]²⁻ 1380.7379, found 1380.7246, calcd for C₁₀₄H₁₉₇B₁₂N₂₅O₅₂+Na [M + Na]⁻ 2782.4663, found 2782.3956.

Synthesis of Carbamate 5c. Clesomer 5c was prepared from 4 (0.10 g, 0.03 mmol) and *tert*-butyl (2-(2-(2-aminoethoxy)ethoxy)-ethyl) carbamate (0.50 g, 2.00 mmol) in ACN (15 mL) using the aforementioned general procedure for carbamate synthesis. The product was obtained as a colorless oil. Yield: 105 mg (84%). IR (neat): 3350, 3012, 2979, 2930, 2108, 1701, 1507, 1252, 1216, and 757 cm⁻¹. ¹H NMR (400 MHz, CD₃CN): δ 6.53 (bs, 11H, NH), 5.67 (bs, 11H, NH), 4.08 (m, 2H, -CH₂-PEG), 3.66–3.47 (m, 118H, -CH₂-PEG), 3.39 (t, 2H, *J* = 5.2 Hz, -CH₂-PEG), 3.22 (m, 42H, -CH₂-PEG), 1.43 (s, 99H, -Boc). ¹³C NMR (100.6 MHz, CD₃CN): δ 157.1, 79.8, 71.3–70.2 (multiple peaks), 51.6, 42.10, 41.1, 28.6. ¹¹B NMR (160.4 MHz, CD₃CN): δ -14.62, -18.15. HRMS (*m/z*): calcd for C₁₄₈H₂₈₅B₁₂N₂₅O₇₄ [M]²⁻ 1864.5215, found 1864.4845.

Synthesis of Chloroacetate Clesomer 6. A solution of 3 (0.43 g, 0.36 mmol) and chloroacetic anhydride (2.70 g, 15.8 mmol) in ACN (30 mL) was refluxed for 3 days in an argon atmosphere with vigorous stirring. Progress of the reaction was monitored by mass spectrometry analysis. The reaction mixture was then concentrated to dryness and purified using a size-exclusion column (Lipophilic Sephadex LH-20) with ACN as the eluent. The product was obtained as a light brown semisolid. Yield: 0.63 g (86%). IR (neat): 2962, 2876, 2107, 1751, 1327, and 1228 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ 4.19–4.11 (m, 22H, -OCOCH₂-), 3.71–3.67 (m, 30H, -CH₂-PEG), 3.41 (t, 2H, *J* = 5.2 Hz, -CH₂-PEG), 3.18 (m, 16H, -CH₂-TBA⁺), 1.66 (m, 16H, -CH₂-TBA⁺), 1.45 (m, 16H, -CH₂-TBA⁺), 1.04 (t, *J* = 4.0 Hz, 24H, CH₃-TBA⁺). ¹³C NMR (100.6 MHz, CDCl₃): δ 166.2, 71.5–70.9 (multiple peaks), 59.7, 51.5, 44.3, 24.8, 20.5, 14.4. ¹¹B NMR (160.4 MHz, CD₃CN): δ -14.61, -18.17. HRMS (*m/z*): calcd for C₃₈H₅₄B₁₂Cl₁₁N₃O₃₀ [M]²⁻ 776.0251, found 776.0341, calcd for C₃₈H₅₄B₁₂Cl₁₁N₃O₃₀ + Na⁺ [M + Na]⁻ 1575.0405, found 1575.0507.

Synthesis of Chloroacetate Clesomer 7. Clesomer 6 (0.48 g, 0.24 mmol), *tert*-butyl prop-2-yn-1-ylcarbamate (0.07 g, 0.47 mmol), and copper iodide (0.05 g, 0.24 mmol) were dissolved in a 50:50 mixture of tetrahydrofuran (THF) and ACN (12 mL). To this mixture was added DIPEA (0.06 g, 0.47 mmol), and the reaction mixture was vigorously stirred at room temperature for 12 h under an argon atmosphere. After completion, the reaction mixture was concentrated to dryness, dissolved in ethyl acetate, and filtered through a Celite pad. The filtrate was concentrated and purification via size-exclusion column chromatography (lipophilic Sephadex LH-20) using ACN afforded the pure product as a colorless oil. Yield: 505 mg (98%). IR (neat): 2963, 2876, 1750, 1328, and 1227 cm⁻¹. ¹H NMR (400 MHz, CD₃CN): δ 7.77 (s, 1H, -CH-triazole), 5.85 (bs, 1H, NH), 4.50 (t, 2H, *J* = 5.2 Hz, -CH₂-NH(Boc)), 4.31 (d, 2H, *J* = 6 Hz, -NCH₂-PEG), 4.20–4.10 (m, 10H, -OCOCH₂Cl), 4.06–3.99 (m, 10H, -OCOCH₂Cl), 3.95 (m, 2H, -OCOCH₂Cl), 3.86 (t, 2H, *J* = 5.2 Hz, -CH₂-PEG), 3.61–3.53 (m, 26H, -CH₂-PEG), 3.48 (t, 2H, *J* = 5.2 Hz, -CH₂-PEG), 3.10 (m, 12H, -CH₂-TBA⁺), 1.62 (m, 12H, -CH₂-TBA⁺), 1.45 (s, 9H, Boc), 1.36 (m, 12H, -CH₂-TBA⁺), 0.99 (t, *J* = 7.2

Hz, 17H, $\text{CH}_3\text{-TBA}^+$) (lesser proton counts for TBA^+ is due to the exchange with H_3O^+ and Na^+). ^{13}C NMR (100.6 MHz, CD_3CN): δ 166.1–165.9 (multiple peaks), 146.7, 123.8, 79.4, 73.3, 71.2–70.8 (multiple peaks), 69.8, 66.4, 59.2, 59.2, 50.8, 43.9, 43.8, 36.7, 28.5, 24.1, 20.2, 13.6. ^{11}B NMR (160.4 MHz, CD_3CN): δ -14.38, -18.12. HRMS (m/z): calcd for $\text{C}_{46}\text{H}_{67}\text{B}_{12}\text{Cl}_{11}\text{N}_4\text{O}_{32} + \text{Na}^+ [\text{M} + \text{Na}]^-$ 1730.1356, found 1730.1433, calcd for $\text{C}_{46}\text{H}_{67}\text{B}_{12}\text{C}_{111}\text{N}_4\text{O}_{32} [\text{M}]^{2-}$ 853.5726, found 853.5412.

Synthesis of Azidoacetate Closures 8. In a 100 mL round-bottom flask, 7 (0.48 g, 0.22 mmol) and sodium azide (1.57 g, 24.09 mmol) were mixed with 20 mL of dry DMF. This mixture was vigorously stirred at 50 °C for 2 days under an argon atmosphere. The progress of the reaction was monitored by mass spectrometry analysis. After completion, the reaction mixture was filtered through a Celite pad, and the filtrate was concentrated to dryness. The residue was dissolved in ethyl acetate and filtered again through a Celite pad. The filtrate was then collected, evaporated to dryness, and purified using size-exclusion column chromatography (lipophilic Sephadex LH-20) with MeOH as eluent. The product was obtained as a light brown semisolid. Yield: 0.46 g (93%). IR (neat): 2963, 2876, 2106, 1739, 1359, and 1243 cm^{-1} . ^1H NMR (400 MHz, CD_3CN): δ 7.74 (s, 1H, $-\text{CH}$ -triazole), 5.58 (bs, 1H, NH), 4.49 (t, 2H, $J = 5.0$ Hz, $-\text{CH}_2\text{-NHBoc}$), 4.30 (d, 2H, $J = 6$ Hz, $-\text{NCH}_2\text{-PEG}$), 3.95 (m, 2H, $-\text{OCOCH}_2\text{N}_3$), 3.88–3.82 (m, 4H, $-\text{OCOCH}_2\text{N}_3$ and $-\text{CH}_2\text{-PEG}$), 3.79 (m, 8H, $-\text{OCOCH}_2\text{N}_3$), 3.72–3.68 (m, 10H, $-\text{OCOCH}_2\text{N}_3$), 3.63–3.47 (m, 28H, $-\text{CH}_2\text{-PEG}$), 3.10 (m, 12H, $-\text{CH}_2\text{-TBA}^+$), 1.62 (m, 12H, $-\text{CH}_2\text{-TBA}^+$), 1.43 (s, 9H, Boc), 1.38 (m, 12H, $-\text{CH}_2\text{-TBA}^+$), 0.99 (t, 18H, $J = 7.2$ Hz, $\text{CH}_3\text{-TBA}^+$) (lesser proton counts for TBA^+ is due to the exchange with H_3O^+ and Na^+). ^{13}C NMR (100.6 MHz, CD_3CN): δ 167.9–167.4 (multiple peaks), 156.7, 123.6, 79.4, 73.2, 71.0–70.8 (multiple peaks), 69.9, 66.2, 59.2, 52.1, 52.0, 50.7, 36.8, 28.5, 24.2, 20.2, 13.7. ^{11}B NMR (160.4 MHz, CD_3CN): δ -14.53, -18.08. HRMS (m/z): calcd for $\text{C}_{46}\text{H}_{67}\text{B}_{12}\text{N}_{37}\text{O}_{32} [\text{M}]^{2-}$ 889.7980, found 889.7671.

General Procedure for the Synthesis of Click Closures 9. In a 25 mL round-bottom flask, the azide-functionalized closure 8 (1 equiv), alkyne (5 equiv per vertex, total 55 equiv), and copper iodide (1 equiv per vertex, total 11 equiv) were dissolved in a 50:50 mixture of THF and ACN (6 mL). To this mixture was added DIPEA (10 equiv per vertex, total 110 equiv), and the reaction mixture was vigorously stirred at room temperature for 3 days under an argon atmosphere. The progress of the reaction was monitored by mass spectrometry analysis. After completion, the reaction mixture was concentrated to dryness, dissolved in ethyl acetate, and filtered through a Celite pad. The filtrate was concentrated and purified via size-exclusion column chromatography (lipophilic Sephadex LH-20) using ACN as an eluent to afford the pure product.

Synthesis of Click Closure 9a. Using the general strategy described above, 9a was synthesized from closure 8 (100 mg, 0.04 mmol), phenylacetylene (247 mg, 2.42 mmol), copper iodide (92.3 mg, 0.48 mmol), and DIPEA (627 mg, 4.80 mmol). Pure product was obtained as a colorless sticky solid. Yield: 116 mg (78%). IR (neat): 2961, 2875, 1743, 1355, and 1255 cm^{-1} . ^1H NMR (400 MHz, CD_3CN): δ 8.04 (m, 6H, $-\text{CH}$ -triazole), 7.94 (m, 6H, $-\text{CH}$ -triazole), 7.87–7.80 (m, 20H, $-\text{Ph}$), 7.37–7.20 (m, 35H, $-\text{Ph}$), 5.90 (bs, 1H, NH), 5.00–4.86 (m, 22H, $-\text{OCOCH}_2\text{-}$), 4.43 (t, 2H, $J = 4.8$ Hz, $-\text{CH}_2\text{-NHBoc}$), 4.28 (m, 2H, $-\text{NCH}_2\text{-PEG}$), 3.77 (t, 2H, $J = 4.8$ Hz, $-\text{NCH}_2\text{-PEG}$), 3.51–3.38 (m, 28H, $-\text{CH}_2\text{-PEG}$), 3.07 (m, 11H, $-\text{CH}_2\text{-TBA}^+$), 1.59 (m, 11H, $-\text{CH}_2\text{-TBA}^+$), 1.40 (s, 9H, Boc), 1.37–1.30 (m, 11H, $-\text{CH}_2\text{-TBA}^+$), 0.98 (t, 17H, $J = 7.2$ Hz, $\text{CH}_3\text{-TBA}^+$) (lesser proton counts for TBA^+ is due to the exchange with H_3O^+ and Na^+). ^{13}C NMR (100.6 MHz, CD_3CN): δ 166.6, 147.8, 131.7, 129.7, 128.7, 126.4, 123.0, 122.9, 73.2, 70.8–69.7 (multiple peaks), 67.2, 65.7, 66.5, 59.2, 55.1, 53.0, 50.8, 43.3, 40.7, 36.6, 28.5, 24.1, 20.1, 13.6, 12.8. ^{11}B NMR (160.4 MHz, CD_3CN): δ -14.44, -18.22. HRMS (m/z): calcd for $\text{C}_{134}\text{H}_{133}\text{B}_{12}\text{N}_{37}\text{O}_{32} [\text{M}]^{2-}$ 1451.5574, found 1451.5630.

Synthesis of Click Closure 9b. Using the general strategy described above, 9b was synthesized from closure 8 (100 mg, 0.04 mmol), 1-hexyne (200 mg, 2.42 mmol), copper iodide (92.3 mg, 0.48 mmol), and DIPEA (627 mg, 4.80 mmol). Pure product was obtained

as a colorless viscous oil. Yield: 120.0 mg (86%). IR (neat): 2962, 2875, 1741, 1354, and 1254 cm^{-1} . ^1H NMR (400 MHz, CD_3CN): δ 7.62–7.40 (m, 12H, $-\text{CH}$ -triazole), 6.05 (bs, 1H, NH), 4.99–4.83 (m, 22H, $-\text{OCOCH}_2\text{-}$), 4.49 (m, 2H, $-\text{CH}_2\text{-NHBoc}$), 4.30 (m, 2H, $-\text{NCH}_2\text{-PEG}$), 3.84 (t, 4H, $-\text{NCH}_2\text{-PEG}$), 3.63–3.48 (m, 26H, $-\text{CH}_2\text{-PEG}$), 3.11 (m, 4.8H, $-\text{CH}_2\text{-TBA}^+$), 2.67 (m, 22H, $-\text{CH}_2\text{-hexyne}$), 1.66–1.58 (m, 26H, $-\text{CH}_2\text{-hexyne}$, $-\text{CH}_2\text{-TBA}^+$), 1.47–1.31 (m, 40H, Boc, $-\text{CH}_2\text{-hexyne}$, $-\text{CH}_2\text{-TBA}^+$), 0.99 (t, 7H, $J = 7.2$ Hz, $\text{CH}_3\text{-TBA}^+$), 0.96–0.95 (m, 33H, $-\text{CH}_2\text{-hexyne}$) (lesser proton counts for TBA^+ are due to the exchange with H_3O^+ and Na^+). ^{13}C NMR (100.6 MHz, CD_3CN): δ 166.4, 148.8, 124.0, 79.50, 79.1, 78.8, 78.5, 73.2, 71.0–70.8 (multiple peaks), 69.7, 59.2, 55.1, 52.9, 51.0, 32.3, 28.6, 25.9, 24.2, 22.9, 20.2, 14.0, 13.7. ^{11}B NMR (160.4 MHz, CD_3CN): δ -14.43, -18.21. HRMS (m/z): calcd for $\text{C}_{112}\text{H}_{177}\text{B}_{12}\text{N}_{37}\text{O}_{32} [\text{M}]^{2-}$ 1341.7291, found 1341.7576.

Synthesis of Click Closure 9c. Using the general strategy described above, 9c was synthesized from closure 8 (100 mg, 0.04 mmol), propargyl acetate (238 mg, 2.42 mmol), copper iodide (92.3 mg, 0.48 mmol), and DIPEA (627 mg, 4.80 mmol). Pure product was obtained as a colorless viscous oil. Yield: 120 mg (81%). IR (neat): 2961, 2876, 1742, 1354, and 1253 cm^{-1} . ^1H NMR (400 MHz, CD_3CN): δ 7.82–7.71 (m, 12H, $-\text{CH}$ -triazole), 6.02 (bs, 1H, NH), 5.17–5.15 (m, 22H, $-\text{CH}_2\text{-OAc}$), 5.09–4.91 (m, 22H, $-\text{OCOCH}_2\text{-}$), 4.49 (t, 2H, $J = 4.4$ Hz, $-\text{CH}_2\text{-NHBoc}$), 4.31 (m, 2H, $-\text{NCH}_2\text{-PEG}$), 3.84 (t, 2H, $J = 4.8$ Hz, $-\text{NCH}_2\text{-PEG}$), 3.75 (m, 2H, $-\text{CH}_2\text{-PEG}$), 3.56–3.47 (m, 26H, $-\text{CH}_2\text{-PEG}$), 3.11 (m, 6H, $-\text{CH}_2\text{-TBA}^+$), 2.00–1.94 (m, 33 H, $-\text{OAc}$), 1.63 (m, 6H, $-\text{CH}_2\text{-TBA}^+$), 1.41 (s, 9H, Boc), 1.36 (m, 6H, $-\text{CH}_2\text{-TBA}^+$), 0.98 (t, 8H, $J = 7.2$ Hz, $\text{CH}_3\text{-TBA}^+$) (lesser proton counts for TBA^+ is due to the exchange with H_3O^+ and Na^+). ^{13}C NMR (100.6 MHz, CD_3CN): δ 171.3, 166.5, 143.4, 126.8, 79.5, 73.1, 70.9–70.5 (multiple peaks), 69.7, 66.5, 59.2, 58.1, 55.8, 52.9, 52.9, 51.0, 43.8, 36.6, 28.5, 24.1, 20.2, 18.6, 17.3, 13.6, 12.8. ^{11}B NMR (160.4 MHz, CD_3CN): δ -14.44, -18.21. HRMS (m/z): calcd for $\text{C}_{101}\text{H}_{133}\text{B}_{12}\text{N}_{37}\text{O}_{54} [\text{M}]^{2-}$ 1429.0017, found 1428.9999.

■ ASSOCIATED CONTENT

Supporting Information

^1H , ^{13}C , and ^{11}B NMR and HRMS spectra of compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

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